

Schwender, Holger; Ickstadt, Katja

**Working Paper**

## Imputing missing genotypes with weighted k nearest neighbors

Technical Report, No. 2008,03

**Provided in Cooperation with:**

Collaborative Research Center 'Reduction of Complexity in Multivariate Data Structures' (SFB 475),  
University of Dortmund

*Suggested Citation:* Schwender, Holger; Ickstadt, Katja (2008) : Imputing missing genotypes with weighted k nearest neighbors, Technical Report, No. 2008,03, Technische Universität Dortmund, Sonderforschungsbereich 475 - Komplexitätsreduktion in Multivariaten Datenstrukturen, Dortmund

This Version is available at:

<https://hdl.handle.net/10419/36594>

**Standard-Nutzungsbedingungen:**

Die Dokumente auf EconStor dürfen zu eigenen wissenschaftlichen Zwecken und zum Privatgebrauch gespeichert und kopiert werden.

Sie dürfen die Dokumente nicht für öffentliche oder kommerzielle Zwecke vervielfältigen, öffentlich ausstellen, öffentlich zugänglich machen, vertreiben oder anderweitig nutzen.

Sofern die Verfasser die Dokumente unter Open-Content-Lizenzen (insbesondere CC-Lizenzen) zur Verfügung gestellt haben sollten, gelten abweichend von diesen Nutzungsbedingungen die in der dort genannten Lizenz gewährten Nutzungsrechte.

**Terms of use:**

*Documents in EconStor may be saved and copied for your personal and scholarly purposes.*

*You are not to copy documents for public or commercial purposes, to exhibit the documents publicly, to make them publicly available on the internet, or to distribute or otherwise use the documents in public.*

*If the documents have been made available under an Open Content Licence (especially Creative Commons Licences), you may exercise further usage rights as specified in the indicated licence.*

# Imputing Missing Genotypes with Weighted $k$ Nearest Neighbors

Holger Schwender\*, Katja Ickstadt

Collaborative Research Center 475, Faculty of Statistics, Dortmund University of Technology, Dortmund, Germany

## Abstract

**Motivation:** Missing values are a common problem in genetic association studies concerned with single nucleotide polymorphisms (SNPs). Since most statistical methods cannot handle missing values, they have to be removed prior to the actual analysis. Considering only complete observations, however, often leads to an immense loss of information. Therefore, procedures are needed that can be used to replace such missing values. In this article, we propose a method based on weighted  $k$  nearest neighbors that can be employed for imputing such missing genotypes.

**Results:** In a comparison to other imputation approaches, our procedure called KNNcatImpute shows the lowest rates of falsely imputed genotypes when applied to the SNP data from the GENICA study, a study dedicated to the identification of genetic and gene-environment interactions associated with sporadic breast cancer. Moreover, in contrast to other imputation methods that take all variables into account when replacing missing values of a particular variable, KNNcatImpute is not restricted to association studies comprising several ten to a few hundred SNPs, but can also be applied to data from whole-genome studies, as an application to a subset of the HapMap data shows.

**Availability:** KNNcatImpute is implemented in the R package `scime` that can be downloaded from <http://cran.r-project.org>.

**Contact:** [holger.schwender@udo.edu](mailto:holger.schwender@udo.edu)

## 1 Introduction

Variations in the human genome are assumed to play an important role in the development of diseases. If such a genetic variation occurs at a single base pair position and in at least 1% of the population, it is called a single nucleotide polymorphism (SNP). Typically, SNPs are biallelic, i.e. two alternatives exist at the corresponding base pair position. Therefore, such SNPs can take three forms: If the base on each of the two chromosomes is of the more/less frequent variant, then a SNP is of the homozygous reference/variant genotype. If one base is of the more frequent, and the other of the less frequent variant, then the SNP is of the heterozygous genotype.

A major goal of genetic association studies is the identification of SNPs and – more importantly – interactions of SNPs (Culverhouse *et al.*, 2002; Garte, 2001). Several methods have

---

\*to whom correspondence should be addressed

been proposed for the detection of SNP interactions. They reach from exhaustive searches based on, e.g., multiple testing (Marchini *et al.*, 2005; Goodman *et al.*, 2006; Ritchie *et al.*, 2001) over the use of evolutionary algorithms (Nunkesser *et al.*, 2007) to approaches based on discrimination procedures (Lunetta *et al.*, 2004; Kooperberg and Ruczinski, 2005; Schwender and Ickstadt, 2007) such as Random Forests (Breiman, 2001) and logic regression (Ruczinski *et al.*, 2003). In Heidema *et al.* (2006) and Hoh and Ott (2003), overviews on such methods are presented.

A problem frequently occurring in association studies is that for some of the observations the genotypes of some of the SNPs are not available. Since most discrimination and feature selection procedures cannot handle missing values, they have to be removed prior to the application of these methods.

The most consistent solution to this problem would be to consider complete observations only, i.e. to remove all observations with one or more missing values. This, however, often leads to an immense loss of information. Moreover, this approach might bias the results of further analyses (Greenland and Finkle, 1995). For example, in the GENICA study (see Section 3), a study dedicated to the identification of genetic and gene-environment interactions associated with sporadic breast cancer, using complete observations would mean that data of just 63.3% of the women are considered (see Figure 1). Therefore, procedures are required that enable the replacement of missing values instead of removing them and the corresponding observations.

In the analysis of gene expression data, KNNimpute proposed by Troyanskaya *et al.* (2002) is a popular method to impute missing expression values based on weighted  $k$  nearest neighbors ( $k$ NN; Fix and Hodges, 1951). Let's assume that the expression value  $x_{ij}$  of gene  $i$  and observation  $j$  is missing, and that  $\mathcal{L}_k$  is a set comprising the  $k$  genes showing the smallest Euclidean distance to gene  $i$  and having a value present for the  $j$ th sample. Using KNNimpute,  $x_{ij}$  is replaced by

$$x_{ij} = \sum_{\ell \in \mathcal{L}_k} w_{i\ell} x_{\ell j} / \sum_{\ell \in \mathcal{L}_k} w_{i\ell}, \quad (1)$$

where the weight  $w_{i\ell}$  is given by the reciprocal of the Euclidean distance between the expression values of gene  $i$  and gene  $\ell$ .

For an application of KNNimpute to protein expression data and a comparison of the Euclidean distance with other distance measures and of the weighted mean (1) with other estimates of average in the context of KNNimpute, see Jung *et al.* (2006).

However, KNNimpute cannot be employed for replacing missing genotypes, since SNPs are categorical variables. In this article, we therefore introduce an approach based on weighted  $k$  nearest neighbors enabling the imputation of missing categorical data. This procedure called KNNcatImpute is then compared to other imputation methods such as the tree-based procedure proposed by Dai *et al.* (2006) and the imputation approach of Random Forests (Breiman, 2001) in their applications to the SNP data from the GENICA study.

Contrary to other non ad hoc imputation methods that take the information in all variables into account when replacing missing values, KNNcatImpute is not restricted to "classical" association studies in which several ten to a few hundred SNPs are considered, but can also be applied to data from whole-genome studies. To exemplify this, we employ this approach to impute the missing genotypes in a subset of the SNP data from the HapMap study (The International HapMap Consortium, 2003) consisting of 262,264 SNPs and 90 persons. Moreover, we compare KNNcatImpute with imputation methods that can also be used for replacing

missing values in such high-dimensional data.

This paper is organized as follows. In Section 2, KNNcatImpute is presented in detail, and it is shown how matrix algebra can be employed to determine all distances required by  $k$  nearest neighbors simultaneously. While in Section 3 this procedure is applied to the GENICA data set and compared with other approaches, Section 4 consists of the comparison of KNNcatImpute and other imputation procedures in their application to the subset of the HapMap data.

## 2 Imputation of missing genotypes based on $k$ nearest neighbors

### 2.1 Distance measures for categorical data

If  $k$  nearest neighbors are to be determined for categorical data, a distance measure has to be employed that can cope with this type of data. Such measures are typically based on an  $R \times C$  contingency table in which the joint distribution of two variables, say  $Y$  and  $Z$ , with observation vectors  $\mathbf{y}$  and  $\mathbf{z}$ , each of length  $n$ , is represented by the numbers

$$n_{rc} = \sum_{j=1}^n I(y_j = r) I(z_j = c) \quad (2)$$

of observations showing the  $r$ th level at  $Y$ ,  $r = 1, \dots, R$ , and the  $c$ th level at  $Z$ ,  $c = 1, \dots, C$ .

An example for such a distance measure is given by

$$d_{\text{Cont}}(\mathbf{y}, \mathbf{z}) = \sqrt{1 - \text{Cont}^2(\mathbf{y}, \mathbf{z})}, \quad (3)$$

where the corrected Pearson's contingency coefficient

$$\text{Cont}(\mathbf{y}, \mathbf{z}) = \sqrt{\frac{\min\{R, C\}}{\min\{R, C\} - 1} \cdot \frac{\chi^2}{\chi^2 + n}}$$

is based on Pearson's  $\chi^2$ -statistic

$$\chi^2 = \sum_{r=1}^R \sum_{c=1}^C \frac{(n_{rc} - \tilde{n}_{rc})^2}{\tilde{n}_{rc}} = \sum_{r=1}^R \sum_{c=1}^C \frac{n_{rc}^2}{\tilde{n}_{rc}} - n$$

for testing the null hypothesis that the two variables are independent by comparing  $n_{rc}$  with

$$\tilde{n}_{rc} = \frac{1}{n} \sum_{c=1}^C n_{rc} \sum_{r=1}^R n_{rc}, \quad (4)$$

i.e. the numbers of observations expected under the null hypothesis.

In this article, we assume that  $R = C$ , i.e. that all variables exhibit the same number of categories. In the case of SNPs,  $R = 3$ , where 1 denotes for the homozygous reference genotype (which is the form that typically shows up most often), 2 the heterozygous genotype, and 3 the homozygous variant.

Besides distances based on Pearson's  $\chi^2$ -statistic for testing independence of two variables, there also exist measures for categorical data that take the number of matches between two

variables into account. A popular example is the distance

$$d_{\text{SMC}}(\mathbf{y}, \mathbf{z}) = 1 - s_{\text{SMC}}(\mathbf{y}, \mathbf{z}) = 1 - \frac{1}{n} \sum_{r=1}^R n_{rr} \quad (5)$$

that is based on the (generalized) simple matching coefficient  $s_{\text{SMC}}$ .

Another measure that also uses the total number of agreements/matches

$$a_R = \sum_{r=1}^R n_{rr},$$

but additionally takes into account the number of agreements that would have been found by chance, i.e.  $e_R = \sum_r \tilde{n}_{rr}$ , is the distance

$$d_{\text{Cohen}}(\mathbf{y}, \mathbf{z}) = 1 - \kappa_C = 1 - \frac{a_R - e_R}{n - e_R} = \frac{n - a_R}{n - e_R}$$

based on Cohen's kappa  $\kappa_C$  (Cohen, 1960).

All these distance measures treat the SNPs as categorical variables, i.e. assume that the distance between the two homozygous genotypes is equal to the distance between each of the homozygous and the heterozygous genotype. A different idea is to take the number of base exchanges into account such that the distance between the homozygous genotypes (two base exchanges) is twice the distance between a homozygous and the heterozygous genotype (one base exchange). Using the abovementioned coding of the genotypes as numeric values, an appropriate measure for this idea is the scaled Manhattan distance

$$d_{\text{Man}}(\mathbf{y}, \mathbf{z}) = \frac{1}{2n} \sum_{i=1}^n |y_i - z_i|. \quad (6)$$

In Section 3.2 and Section 4.2, these four measures are compared in the application of KNNcatImpute to the SNPs from the GENICA and the HapMap study, respectively.

## 2.2 KNNcatImpute

After having computed the values of an appropriate distance measure  $d$  for all pairs of SNPs, a missing genotype  $x_{ij}$  of SNP  $i$  and observation  $j$  is imputed by first identifying the set  $\mathcal{L}_k$  comprising the  $k$  SNPs showing the smallest distances to SNP  $i$  and having a value present for the  $j$ th observation. Then,  $x_{ij}$  is determined by weighted majority voting, i.e. by

$$x_{ij} = \arg \max_{r=1, \dots, R} \sum_{\ell \in \mathcal{L}_k} d^{-1}(\mathbf{x}_\ell, \mathbf{x}_i) I(x_{\ell j} = r), \quad (7)$$

where  $\mathbf{x}_\ell$  is a vector containing the genotypes of SNP  $\ell$  for all observations. Note that in (7) the normalization factor  $\sum_{\ell \in \mathcal{L}_k} d^{-1}(\mathbf{x}_\ell, \mathbf{x}_i)$  is omitted, since – contrary to (1) – the value of  $x_{ij}$  is not affected by this constant. If  $d(\mathbf{x}_\ell, \mathbf{x}_i) = 0$  for any  $\ell \in \mathcal{L}_k$ , then  $x_{ij}$  is replaced by  $x_{\ell j}$ .

## 2.3 Simultaneous computation of contingency tables

Applying KNNcatImpute to a data set composed of  $m$  variables and  $n$  observations requires the calculation of  $m(m-1)/2$  distances. Since computing these values one-by-one can be very

time-consuming, we employ the vectorization and matrix computation functionalities of the statistical software R (R Development Core Team, 2007) to speed up the computation of the values of the distance measures based on contingency tables.

Suppose that  $\mathbf{X}$  is an  $m \times n$  matrix in which each row represents one of the  $m$  variables, and each column corresponds to one of the  $n$  observations. For a start, assume that none of the values are missing. If  $\mathbf{X}^{(r)}$  denotes an  $m \times n$  indicator matrix,  $r = 1, \dots, R$ , with elements

$$x_{ij}^{(r)} = \begin{cases} 1, & \text{if } x_{ij} = r \\ 0, & \text{if } x_{ij} \neq r \text{ (or } x_{ij} \text{ is missing)} \end{cases},$$

then the upper triangle of the  $m \times m$  matrix

$$\mathbf{N}^{(rc)} = \mathbf{X}^{(r)} \mathbf{X}^{(c)'} \quad (8)$$

contains the numbers  $n_{rc}$  for all  $m(m-1)/2$  pairs of variables, cf. (2), and the lower triangle consists of the numbers  $n_{cr}$  of these pairs,  $r, c = 1, \dots, R$ . More precisely, the value of (2) for the two SNPs represented by the  $i$ th and the  $\ell$ th row of  $\mathbf{X}$  is given by the  $(i\text{th}, \ell\text{th})$  element of  $\mathbf{N}^{(rc)}$ . Analogously,

$$\tilde{\mathbf{N}}^{(rc)} = \frac{1}{n} \mathbf{X}^{(r)} \mathbf{1}_n \mathbf{1}_n' \mathbf{X}^{(c)'} \quad (9)$$

is composed of the expected numbers  $\tilde{n}_{rc}$  and  $\tilde{n}_{cr}$ , cf. (4), where  $\mathbf{1}_n$  denotes a vector of length  $n$  consisting only of ones.

Using (8), the  $m \times m$  distance matrix  $\mathbf{D}_{\text{SMC}}$  containing the distances  $d_{\text{SMC}}$ , see (5), for all  $m(m-1)/2$  pairs of SNPs can be computed by

$$\mathbf{D}_{\text{SMC}} = \mathbf{1}_{m,m} - \frac{1}{n} \sum_{r=1}^R \mathbf{N}_{rr}, \quad (10)$$

where  $\mathbf{1}_{m,m}$  is an  $m \times m$  matrix consisting only of ones. Similarly, the distance matrix  $\mathbf{D}_{\text{Cohen}}$  can be computed by employing (8) and (9).

For the distance matrix  $\mathbf{D}_{\text{Cont}}$ , an  $m \times m$  matrix  $\mathbf{Q}$  containing all  $m(m-1)/2$  values of Pearson's  $\chi^2$ -statistic needs to be determined by

$$\mathbf{Q} = \sum_{r=1}^R \sum_{c=1}^R \frac{\mathbf{N}^{(rc)} * \mathbf{N}^{(rc)}}{\tilde{\mathbf{N}}^{(rc)}} - n, \quad (11)$$

where  $*$  and the fraction line denote elementwise matrix calculations.

The  $m \times m$  distance matrix  $\mathbf{D}_{\text{Man}}$  composed of the values of the scaled Manhattan distance (6) can also be determined by matrix computation. For this, note that if  $y_i$  and  $z_i$ ,  $i = 1, \dots, n$ , are integers between 1 and  $R$ , then (6) can also be calculated by

$$d_{\text{Man}}(\mathbf{y}, \mathbf{z}) = \frac{1}{2n} \sum_{r=1}^R \sum_{\substack{c=1 \\ c \neq r}}^R |r - c| \cdot n_{rc}$$

such that  $\mathbf{D}_{\text{Man}}$  is given by

$$\mathbf{D}_{\text{Man}} = \frac{1}{2n} \sum_{r=1}^R \sum_{\substack{c=1 \\ c \neq r}}^R |r - c| \cdot \mathbf{N}^{(rc)}. \quad (12)$$

**TABLE 1.** Processing times (in seconds) of both the matrix algebra based and the individual calculation of the values of Pearson’s  $\chi^2$ -statistic for all pairs of  $m$  categorical variables, where each variable exhibits  $r = 3$  levels, and the number of observations is  $n = 1,000$ .

| $m$        | 10   | 50   | 100   | 200   | 500    |
|------------|------|------|-------|-------|--------|
| Matrix     | 0.01 | 0.07 | 0.33  | 1.22  | 7.79   |
| Individual | 0.15 | 4.25 | 17.32 | 70.06 | 474.22 |

If there are missing values in  $\mathbf{X}$ , the matrix algebra based procedure will lead to incorrect distances for variables exhibiting missing values, since the number of observations showing no missing value at such a variable will differ from the total number  $n$  of observations. To extend this approach to a matrix  $\mathbf{X}$  containing missing values, it is therefore necessary to define an  $m \times n$  matrix  $\mathbf{X}^{\text{NA}}$  with elements

$$x_{ij}^{\text{NA}} = \begin{cases} 1, & \text{if } x_{ij} \text{ is not missing} \\ 0, & \text{if } x_{ij} \text{ is missing} \end{cases},$$

and to replace  $n$  in (9)-(12) by the  $m \times m$  matrix  $\mathbf{N} = \mathbf{X}^{\text{NA}} (\mathbf{X}^{\text{NA}})'$ .

Since the rowwise sums of  $\mathbf{X}^{(r)}$  in (9) only take individual but not pairwise missing values, i.e. missing values appearing in either of the two considered variables, into account, it additionally is necessary to replace  $\mathbf{X}^{(r)} \mathbf{1}_n$  by  $\mathbf{Z}^{(r)} = \mathbf{X}^{(r)} (\mathbf{X}^{\text{NA}})'$  such that (9) becomes

$$\tilde{\mathbf{N}}^{(rc)} = \frac{\mathbf{Z}^{(r)} (\mathbf{Z}^{(c)})'}{\mathbf{N}}.$$

## 2.4 Comparison of processing times

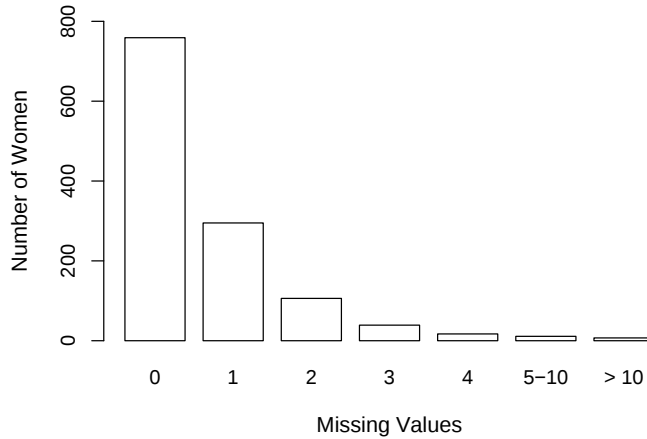
To evaluate how much an implementation of the procedure based on matrix calculation in R accelerates the computation in comparison to an individual determination of the  $m(m-1)/2$  values of Pearson’s  $\chi^2$ -statistic, both approaches are applied to several numbers  $m$  of variables.

In Table 1, the resulting processing times on an AMD Athlon XP 3000+ machine with 1 GB of RAM are summarized. This table reveals that using the vectorization and matrix computation functionalities of R speeds up computation substantially, in particular if  $m$  is large. If, e.g., 500 variables are considered, then it takes just less than 8 seconds to obtain the 124,750 values of Pearson’s  $\chi^2$ -statistics when employing matrix computation, whereas an individual calculation requires about 8 minutes.

## 3 Application to an Association Study

### 3.1 GENICA data set

The GENICA study is a case-control study carried out by the interdisciplinary study group on **G**ene **EN**vironment Interaction and breast **C**ancer in Germany (<http://www.genica.de>), a joint initiative of several German research institutes dedicated to the detection of genetic and environmental factors leading to a higher risk of developing sporadic breast cancer. The cases and controls of this age-matched and population-based study have been recruited in the



**FIGURE 1.** Distribution of the numbers of missing values over the 1,234 observations from the GENICA study.

larger Bonn region, Germany, between 2000 and 2002. For more details, see Justenhoven *et al.* (2004).

In this article, the focus is on a subset of the genotype data from the GENICA study. More precisely, data of 1,234 women (609 cases and 625 controls) and 39 SNPs belonging to the estrogen, the DNA repair, or the control of cell cycle pathway are available for the analysis.

Since a few of the women show many missing genotypes (see Figure 1), all 35 observations with more than three missing values are removed from the analysis leading to a total of 1,199 women (592 cases and 607 controls). The missing values of these women could, of course, also have been imputed. We, however, prefer to remove such observations – as long as their number is small – since observations with many replaced values may add large uncertainties to further analyses with, e.g., discrimination methods.

The remaining 1.3% missing values should be replaced by the imputation method that performs best in a comparison of already existing approaches for imputing categorical data with KNNcatImpute. But first we would like to investigate which parameter setting is best suited for an application of KNNcatImpute to the GENICA data set.

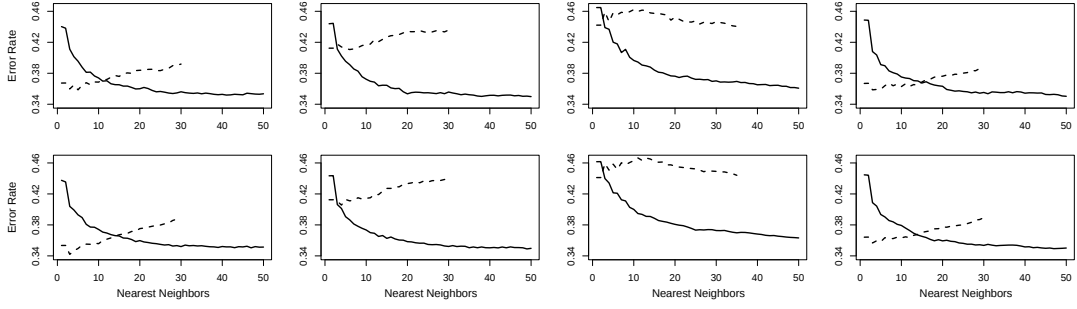
### 3.2 Comparison of parameter settings

Contrary to the discrimination approach of  $k$  Nearest Neighbors (see., e.g., Ripley, 1996) in which the  $k$  nearest observations are employed to predict the class of a new observation, Troyanskaya *et al.* (2002) borrow strength from a huge number of genes by imputing the missing values based on the  $k$  nearest genes.

In the GENICA data set, however, the number of observations is much larger than the number of SNPs. To figure out which approach works best for the GENICA data, KNNcatImpute therefore is not only applied to  $\mathbf{X}$  to identify the  $k$  nearest SNPs, but also to  $\mathbf{X}'$  to detect the  $k$  nearest observations.

Moreover, it is examined which of the distance measures presented in Section 2.1 is most suitable in the analysis of the GENICA data set.

For these comparisons, only the genotypes of the 759 complete observations, i.e. women without missing values, are taken into account. Since in the GENICA data set 1.3% of the



**FIGURE 2.** Mean fractions of falsely imputed values if  $d_{\text{SMC}}$  (left-most panel),  $d_{\text{Cohen}}$  (second left-most panel),  $d_{\text{Cont}}$  (second right-most panel), or  $d_{\text{Man}}$  (right-most panel) is employed in KNNcatImpute to replace the 1% (upper panel) or 2% (lower panel) artificially generated missing values in the GENICA data set. The solid lines mark the error rates when  $k$  nearest observations are used, and the dashed lines the error rates when  $k$  nearest SNPs are considered.

genotypes are missing and the common rate of missing values in association studies is 5%-10% (Dai *et al.*, 2006), we remove randomly 1%, 2%, 5%, and 10% of the genotypes, respectively. Afterwards, the (artificial) missing values are replaced using the different settings of KNNcatImpute, and the imputed values are compared with the real genotypes. This procedure is repeated 50 times.

In Figure 2, the mean fractions of falsely imputed genotypes for different values of  $k$  and different settings of KNNcatImpute are displayed. (Since for all four cases, i.e. 1%, 2%, 5%, and 10% artificially removed values, the plots look similar, only the error rates for the imputations of the 1% and 2% missing values are presented.) This figure reveals that the distance  $d_{\text{Cont}}$  based on the corrected Pearson’s contingency coefficient exhibits larger error rates than the three other measures when used in KNNcatImpute. The other three measures perform almost equally well when searching for the  $k$  nearest observations, whereas  $d_{\text{Cohen}}$  shows larger error rates than  $d_{\text{SMC}}$  and  $d_{\text{Man}}$  when considering the  $k$  nearest SNPs. For small values of  $k$ ,  $k$  nearest SNPs performs better than  $k$  nearest observations which might be due to the fact that in these cases mostly SNPs from the same gene are used to impute the missing genotypes (see also Müller *et al.*, 2005).

Therefore, the most suitable setting of KNNcatimpute for imputing the 1.3% missing genotypes in the GENICA data set seems to be an approach based on  $k$  nearest observations with  $k \approx 50$  and on one of the distance measures  $d_{\text{SMC}}$ ,  $d_{\text{Cohen}}$  and  $d_{\text{Man}}$ .

### 3.3 Comparison of imputation procedures

To determine how well KNNcatImpute performs in comparison to other imputation methods, three ad hoc approaches and two more sophisticated procedures are also applied to the data sets with the artificially generated missing values described in Section 3.2. In this comparison, we do not consider haplotype-based imputation methods (e.g., Dai *et al.*, 2006), since we do not have haplotype information on the SNPs from the GENICA study.

In the three ad hoc approaches, the removed genotypes are imputed by

- the SNP-wise mode, i.e. typically the homozygous reference genotype,
- a random draw from the distribution of the respective SNP,

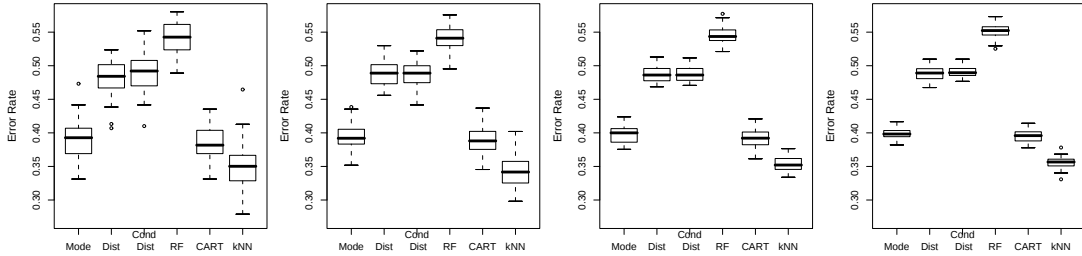
- a random draw from the conditional distribution of the SNP given the case-control status.

The first more sophisticated method is based on Random Forests (Breiman, 2001). In this procedure, the missing values are initially replaced by the mode of the respective variable. Afterwards, Random Forests is applied to this data set, and the proximity, i.e. the fraction of trees containing two particular observations in the same terminal node, is computed for each pair of observations. The missing genotypes are then recalculated by weighted majority voting, where the votes resulting from the trees are weighted by the proximities. This procedure is repeated several times, and the values determined in the final iteration are the estimates for the missing genotypes.

The second approach proposed by Dai *et al.* (2006) is based on a combination of Gibbs sampling (see, e.g., Gelman *et al.*, 2003) and CART (Breiman *et al.*, 1984). Gibbs sampling is used to iteratively sample from the conditional distribution of the missing data of the  $j$ th observation given the values computed for the missing genotypes of the other observations in the previous steps of Gibbs sampling and the complete data of all observations, whereas CART is employed to model this full conditional distribution (for more details, see Dai *et al.*, 2006).

For the comparison, the settings of KNNcatImpute leading to the best results in Section 3.2 are used, whereas the two more sophisticated methods are optimized by considering different values of their parameters. While in the CART based approach 1, 2, 5, 10 (recommended by Dai *et al.*, 2006), 15 and 20 iterations are examined, the Random Forests based method is applied to the data sets using, on the one hand, one to six iterations, and on the other hand, 500, 1000 and 2000 trees with  $\lfloor \sqrt{39} \rfloor = 6, 3, 12$  and 39 SNPs selected randomly from the 39 GENICA SNPs at each node (cf. Breiman, 2001).

In Figure 3, the fractions of falsely imputed genotypes of the applications of these procedures with optimized parameters are displayed. This figure reveals that KNNcatImpute leads to lower error rates than the approach of Dai *et al.* (2006) and the imputations based on the mode which in turn exhibit lower error rates than the other procedures. Therefore, KNNcatImpute seems to be best suited for the imputation of the missing values from the GENICA data set, and is therefore used to replace these genotypes.



**FIGURE 3.** Boxplots of the fractions of falsely imputed genotypes when replacing the 1% (left-most panel), 2% (second left-most panel), 5% (second right-most panel), or 10% (right-most panel) missing values by the mode, by a draw from the SNP-wise distribution, by a draw from the conditional distribution of the SNP given the case-control status, by the Random Forests based method (5 Iterations, 500 trees with 6 SNPs at each node), by the CART-based procedure of Dai *et al.* (2006) with one iteration, and by KNNcatImpute.

## 4 Application to Whole-Genome Data

### 4.1 HapMap data set

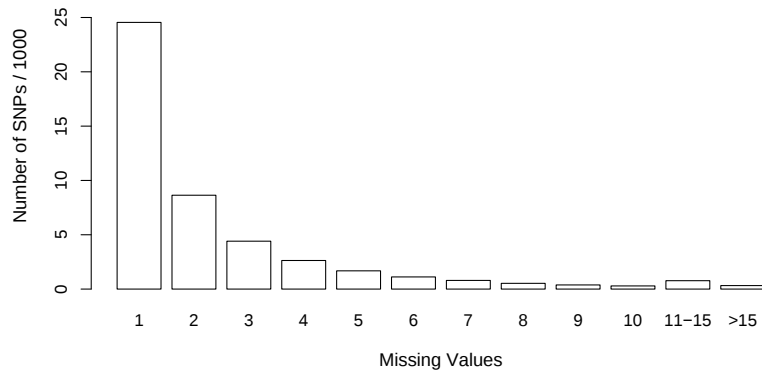
The International HapMap Project (<http://www.hapmap.org>; The International HapMap Consortium, 2003) is a collaboration of several scientific groups from different countries. In this project, millions of SNPs have been genotyped for each of 270 persons from four populations, namely 45 Japanese from Tokyo, 45 Han Chinese from Beijing, 90 Yoruba in Ibadan, Nigeria, and 90 CEPH (Utah residents with ancestry from Northern and Western Europe).

About 500,000 of these SNPs have been measured using the Affymetrix GeneChip Mapping 500K Array Set composed of the Nsp and the Sty array. In this article, we focus on the genotypes of the 262,264 SNPs from the Nsp arrays that have been determined by the standard Affymetrix genotype calling algorithm BRLMM (Bayesian Robust Linear Models with Mahalanobis distance; Affymetrix, 2006). These genotypes can be downloaded from [http://www.affymetrix.com/support/technical/sample\\_data/500k\\_hapmap\\_genotype\\_data.affx](http://www.affymetrix.com/support/technical/sample_data/500k_hapmap_genotype_data.affx).

Furthermore, we only consider the genotypes of the 45 Japanese and 45 Han Chinese, since all these individuals are unrelated, whereas each of the other two populations consists of 30 trios each of which is composed of data from a mother, a father and their child. For imputing missing genotypes in the latter two populations, it therefore might be more appropriate to take into account the relationship between the observations.

Since 45,793 of the 262,264 SNPs are monomorph, i.e. show only one genotype, and therefore useless for discriminating the two populations, they are removed from the data set prior to the application of the imputation methods, although all procedures considered in this section can handle such SNPs. For 46,118 of the remaining 216,471 SNPs, at least one of the 90 genotypes is missing. In Figure 4, the distribution of the numbers of missing values for these 46,118 SNPs is displayed.

As a relatively small number of SNPs exhibits a large number of missing values, all SNPs exhibiting less than 90% non-missing values are removed leading to 44,733 SNPs with at least one missing genotypes. The remaining 0.49% missing values in this subset of the HapMap data set should again be replaced by the imputation method leading to the smallest numbers of falsely imputed values.



**FIGURE 4.** Distribution of the numbers of missing genotypes in the 46,118 SNPs from the HapMap data that show at least one missing value.

**TABLE 2.** Mean fractions of falsely imputed genotypes when applying KNNcatImpute with different distance measures and different numbers of nearest neighbors to the HapMap data sets in which either 1% or 2% of the values have been removed randomly.

|    |         | $k$                | 1     | 3     | 5     | 7     |
|----|---------|--------------------|-------|-------|-------|-------|
| 1% |         | $d_{\text{SMC}}$   | 0.142 | 0.147 | 0.153 | 0.158 |
|    |         | $d_{\text{Cohen}}$ | 0.148 | 0.153 | 0.156 | 0.159 |
|    | Removed | $d_{\text{Cont}}$  | 0.531 | 0.542 | 0.549 | 0.556 |
|    |         | $d_{\text{Man}}$   | 0.140 | 0.145 | 0.150 | 0.155 |
| 2% |         | $d_{\text{SMC}}$   | 0.143 | 0.148 | 0.154 | 0.159 |
|    |         | $d_{\text{Cohen}}$ | 0.150 | 0.154 | 0.157 | 0.160 |
|    | Removed | $d_{\text{Cont}}$  | 0.364 | 0.374 | 0.382 | 0.389 |
|    |         | $d_{\text{Man}}$   | 0.142 | 0.146 | 0.151 | 0.156 |

## 4.2 Comparison of parameter settings

To figure out which of the distance measures and how many nearest neighbors should be used in KNNcatImpute to replace the missing values in the HapMap data set, 1% and 2% of the values of the 170,353 SNPs without missing genotype are removed randomly, and then imputed using KNNcatImpute with different parameter settings to compare the imputed values with the real genotypes. This procedure is repeated 20 times.

Contrary to the application of KNNcatImpute to the GENICA data set in which all SNPs/observations are considered in the search for the  $k$  nearest SNPs/observations, we here “just” use the SNPs without missing values to identify the  $k$  nearest neighbors of a SNP with missing values. Moreover, we restrict the search by considering the SNPs chromosomewise such that the missing genotypes of a particular SNP are imputed using only SNPs that come from the same chromosome as the considered SNP. The latter is not only time-saving, but also biologically meaningful, as only SNPs from the same chromosome are inherited together.

In Table 2, the mean fractions of falsely imputed values are summarized for the different settings of KNNcatImpute. This table shows that while employing the corrected Pearson’s contingency coefficient works poorly also in the application to the HapMap data, the other three distance measures perform almost equally well, where the scaled Manhattan distance exhibits slightly lower error rates than  $d_{\text{SMC}}$  which in turn leads to slightly less falsely imputed genotypes than  $d_{\text{Cohen}}$ . For all distance measures, using one nearest neighbor in KNNcatImpute performs best, whereas employing seven nearest neighbors leads to the largest error rates.

## 4.3 Comparison of imputation procedures

For comparison, the three ad hoc methods from Section 3.3 are also applied to the data sets with artificially generated missing genotypes. The imputation methods of Dai *et al.* (2006) and Breiman (2001), however, cannot be used to replace these missing values, as it is not feasible to apply these procedure to whole-genome data, even if the SNPs with missing genotypes are considered chromosomewise. This shows an advantage of KNNcatImpute: Contrary to other approaches that take not only the values of the variable whose missing values should

**TABLE 3.** Minimum, mean and maximum of the fractions of falsely imputed genotypes determined in applications of four imputation methods to the HapMap data sets in which either 1% or 2% of the values have been removed randomly.

|    |                    | Minimum | Mean  | Maximum |
|----|--------------------|---------|-------|---------|
| 1% | KNNcatImpute       | 0.138   | 0.140 | 0.142   |
|    | Mode               | 0.381   | 0.384 | 0.386   |
|    | Distribution       | 0.441   | 0.443 | 0.445   |
|    | Cond. Distribution | 0.440   | 0.441 | 0.443   |
| 2% | KNNcatImpute       | 0.140   | 0.142 | 0.143   |
|    | Mode               | 0.383   | 0.384 | 0.387   |
|    | Distribution       | 0.442   | 0.443 | 0.445   |
|    | Cond. Distribution | 0.440   | 0.441 | 0.444   |

be replaced into account, but employ the information of all variables to impute these missing values, KNNcatImpute can be applied to whole-genome data.

Table 3 in which the fractions of falsely imputed genotypes of both the three ad hoc methods and KNNcatImpute using  $d_{\text{Man}}$  and  $k = 1$  nearest neighbor are displayed reveals that KNNcatImpute exhibits substantially smaller error rates than the other imputation methods.

We therefore replace the 93,927 missing genotypes in the subset of the HapMap data set described in Section 4.1 by applying KNNcatImpute to these data, which takes about 28 minutes on an AMD Athlon XP 3000+ with 1 GB of RAM.

## 5 Discussion

Missing genotypes are a common problem in association studies. Since many statistical methods cannot handle missing values, they need to be removed or replaced prior to the actual analysis. In this article, we have presented a method based on weighted  $k$  nearest neighbors for imputing missing values in categorical data such as SNP data.

In a comparison with other imputation methods, this procedure called KNNcatImpute shows the lowest fractions of falsely imputed genotypes when applied to the SNP data from the GENICA study.

In our comparison, no haplotype-based imputation approach has been considered, since for the GENICA data set we do not have information on haplotypes, i.e. on blocks of SNPs that are inherited together and are therefore (highly) correlated. If such information is available, haplotype-based methods might improve the imputation of missing genotypes. However, KNNcatImpute is able to identify groups of highly correlated SNPs, and should therefore also work well in comparison to such approaches when haplotype information is available.

An advantage of KNNcatImpute over other imputation methods that also take not only the distribution of a particular SNP, but the information from all SNPs into account to impute the missing values of this SNP is that KNNcatImpute can be applied to data from whole-genome studies, whereas it is infeasible to use other non ad hoc imputation approaches such as the procedure based on Random Forests and the method of Dai *et al.* (2006) to replace missing

values in such high-dimensional data sets.

To exemplify this KNNcatImpute is applied to a subset of the SNP data from the HapMap project. In a comparison with other imputation methods that only consider the distribution of the SNP whose missing values should be replaced, KNNcatImpute shows the by far smallest fractions of falsely imputed genotypes.

As in the application of KNNcatImpute to the GENICA data set, employing the distance measures  $d_{\text{SMC}}$ ,  $d_{\text{Cohen}}$  and  $d_{\text{Man}}$  that take the numbers of matches between the genotypes or the alleles of two SNPs, respectively, into account lead to almost the same fractions of falsely imputed genotypes in the analysis of the HapMap data set, whereas the distance  $d_{\text{Cont}}$  based on the corrected Pearson’s contingency coefficient, which is comparable to a correlation coefficient in the analysis of continuous data, performs poorly in the applications to both data sets.

Although KNNcatImpute has been developed in the context of SNP data, it is not restricted to such data, but can also be applied to other types of categorical data. However, in such an application all variables should show the same number of categories, and the meaning and the order of the categories should be the same for all variables so that distance measures such as  $d_{\text{SMC}}$  and  $d_{\text{Cohen}}$  can be employed appropriately.

## Acknowledgement

The financial support of the Deutsche Forschungsgemeinschaft (SFB 475, “Reduction of Complexity in Multivariate Data Structures”) is gratefully acknowledged. The authors would also like to thank Tina Müller for fruitful discussions on distance measures, Ingo Ruczinski for providing the R function of the imputation method of Dai *et al.* (2006), and all partners within the GENICA network for their cooperation.

## References

- Affymetrix (2006). BRLMM: An improved genotype calling method for the GeneChip Human Mapping 500k array set. Technical report, Affymetrix, Santa Clara, CA.
- Breiman, L. (2001). Random Forests. *Mach. Learn.*, **45**, 5–32.
- Breiman, L., Friedman, J., Olshen, R., and Stone, C. (1984). *Classification and regression trees*. Wadsworth, Belmont, CA.
- Cohen, J. (1960). A coefficient of agreement for nominal scales. *Educ. Psychol. Meas.*, **20**, 27–46.
- Culverhouse, R., Suarez, B., Lin, J., and Reich, T. (2002). A perspective on epistasis: Limits of models displaying no main effect. *Am. J. Hum. Genet.*, **70**, 461–471.
- Dai, J., Ruczinski, I., LeBlanc, M., and Kooperberg, C. (2006). Imputation methods to improve inference in SNP association studies. *Genet. Epidemiol.*, **30**, 690–702.
- Fix, E. and Hodges, J. (1951). Discriminatory analysis – nonparametric discrimination: consistency properties. Technical report, USAF School of Aviation Medicine.

- Garte, S. (2001). Metabolic susceptibility genes as cancer risk factors: Time for a reassessment? *Cancer Epidemiol. Biomarkers Prev.*, **10**, 1233–1237.
- Gelman, A., Carlin, J., Stern, H., and Rubin, D. (2003). *Bayesian data analysis*. Chapman & Hall, London, second edition.
- Goodman, J., Mechanic, L., Luke, B., Ambs, S., Chanock, S., and Harris, C. (2006). Exploring SNP-SNP interactions and colon cancer risk using polymorphism interaction analysis. *Int. J. Cancer*, **118**, 1790–1797.
- Greenland, S. and Finkle, W. (1995). A critical look at methods for handling missing covariates in epidemiologic regression analysis. *Am. J. Epidemiol.*, **142**, 1255–1264.
- Heidema, G., Boer, J., Nagelkerke, N., Mariman, E., van de A, D., and Feskens, E. (2006). The challenge for genetic epidemiologists: how to analyze large numbers of SNPs in relation to complex diseases. *BioMed Genet.*, **7**(23).
- Hoh, J. and Ott, J. (2003). Mathematical multi-locus approaches to localizing complex human trait genes. *Nat. Rev. Genet.*, **4**, 701–709.
- Jung, K., Gannoun, A., Sitek, B., Apostolov, O., Schramm, A., H.E., M., Stühler, K., and Urfer, W. (2006). Statistical evaluation of methods for the analysis of dynamic protein expression data from a tumor study. *REVSTAT*, **4**, 67–80.
- Justenhoven, C., Hamann, U., Pesch, B., Harth, V., Rabstein, S., Baisch, C., Vollmert, C., Illig, T., Ko, Y., Brüning, T., and Brauch, H. (2004). ERCC2 genotypes and a corresponding haplotype are linked with breast cancer risk in a German population. *Cancer Epidemiol. Biomarkers Prev.*, **13**, 2059–2064.
- Kooperberg, C. and Ruczinski, I. (2005). Identifying interacting SNPs using Monte Carlo logic regression. *Genet. Epidemiol.*, **28**, 157–170.
- Lunetta, K., Hayward, L., Segal, J., and van Eerdewegh, P. (2004). Screening large-scale association study data: exploiting interactions using Random Forests. *BMC Genet.*, **10**(32).
- Marchini, J., Donnelly, P., and Cardon, R. (2005). Genome-wide strategies for detecting multiple loci that influence complex diseases. *Nat. Genet.*, **37**, 413–416.
- Müller, T., Selinski, S., and Ickstadt, K. (2005). Cluster analysis: a comparison of different similarity measures for SNP data. Technical report, SFB 475, Department of Statistics, Dortmund University of Technology.
- Nunkesser, R., Bernholt, T., Schwender, H., Ickstadt, K., and Wegener, I. (2007). Detecting high-order interactions of single nucleotide polymorphisms using genetic programming. *Bioinformatics*. doi: 10.1093/bioinformatics/btm522.
- R Development Core Team (2007). *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0.
- Ripley, B. (1996). *Pattern recognition and neural networks*. Cambridge University Press, Cambridge.

- Ritchie, M., Hahn, L., Roodi, N., Bailey, L., Dupont, W., Parl, F., and Moore, J. (2001). Multifactor-dimensionality reduction reveals high-order interactions among estrogen-metabolism genes in sporadic breast cancer. *Am. J. Hum. Genet.*, **69**, 138–147.
- Ruczinski, I., Kooperberg, C., and LeBlanc, M. (2003). Logic regression. *J. Comput. Graph. Stat.*, **12**, 475–511.
- Schwender, H. and Ickstadt, K. (2007). Identification of SNP interactions using logic regression. *Biostat.* doi:10.1093/biostatistics/kxm024.
- The International HapMap Consortium (2003). The International HapMap Project. *Nature*, **426**, 789–796.
- Troyanskaya, O., Cantor, M., Sherlock, G., Brown, P., Hastie, T., Tibshirani, R., Botstein, D., and Altman, R. (2002). Missing value estimation methods for DNA microarrays. *Bioinformatics*, **17**, 520–525.