

Nunkesser, Robin; Bernholt, Thorsten; Schwender, Holger; Ickstadt, Katja;  
Wegener, Ing

## Working Paper

# Detecting high-order interactions of single nucleotide polymorphisms using genetic programming

Technical Report, No. 2007,24

## Provided in Cooperation with:

Collaborative Research Center 'Reduction of Complexity in Multivariate Data Structures' (SFB 475),  
University of Dortmund

*Suggested Citation:* Nunkesser, Robin; Bernholt, Thorsten; Schwender, Holger; Ickstadt, Katja; Wegener, Ing (2007) : Detecting high-order interactions of single nucleotide polymorphisms using genetic programming, Technical Report, No. 2007,24, Universität Dortmund, Sonderforschungsbereich 475 - Komplexitätsreduktion in Multivariaten Datenstrukturen, Dortmund

This Version is available at:

<https://hdl.handle.net/10419/36598>

### Standard-Nutzungsbedingungen:

Die Dokumente auf EconStor dürfen zu eigenen wissenschaftlichen Zwecken und zum Privatgebrauch gespeichert und kopiert werden.

Sie dürfen die Dokumente nicht für öffentliche oder kommerzielle Zwecke vervielfältigen, öffentlich ausstellen, öffentlich zugänglich machen, vertreiben oder anderweitig nutzen.

Sofern die Verfasser die Dokumente unter Open-Content-Lizenzen (insbesondere CC-Lizenzen) zur Verfügung gestellt haben sollten, gelten abweichend von diesen Nutzungsbedingungen die in der dort genannten Lizenz gewährten Nutzungsrechte.

### Terms of use:

*Documents in EconStor may be saved and copied for your personal and scholarly purposes.*

*You are not to copy documents for public or commercial purposes, to exhibit the documents publicly, to make them publicly available on the internet, or to distribute or otherwise use the documents in public.*

*If the documents have been made available under an Open Content Licence (especially Creative Commons Licences), you may exercise further usage rights as specified in the indicated licence.*

# Detecting High-Order Interactions of Single Nucleotide Polymorphisms Using Genetic Programming

Robin Nunkesser<sup>1,2\*</sup>, Thorsten Bernholt<sup>1,2</sup>, Holger Schwender<sup>1,3</sup>,  
Katja Ickstadt<sup>1,3</sup>, Ingo Wegener<sup>1,2</sup>

<sup>1</sup> Collaborative Research Center 475, University of Dortmund, Dortmund, Germany

<sup>2</sup> Department of Computer Science, University of Dortmund, Dortmund, Germany

<sup>3</sup> Department of Statistics, University of Dortmund, Dortmund, Germany

## Abstract

**Motivation:** Not individual single nucleotide polymorphisms (SNPs), but high-order interactions of SNPs are assumed to be responsible for complex diseases such as cancer. Therefore, one of the major goals of genetic association studies concerned with such genotype data is the identification of these high-order interactions. This search is additionally impeded by the fact that these interactions often are only explanatory for a relatively small subgroup of patients. Most of the feature selection methods proposed in the literature, unfortunately, fail at this task, since they can either only identify individual variables or interactions of a low order, or try to find rules that are explanatory for a high percentage of the observations. In this paper, we present a procedure based on genetic programming and multi-valued logic that enables the identification of high-order interactions of categorical variables such as SNPs. This method called GPAS (Genetic Programming for Association Studies) cannot only be used for feature selection, but can also be employed for discrimination.

**Results:** In an application to the genotype data from the GENICA study, an association study concerned with sporadic breast cancer, GPAS is able to identify high-order interactions of SNPs leading to a considerably increased breast cancer risk for different subsets of patients that are not found by other feature selection methods. As an application to a subset of the HapMap data shows, GPAS is not restricted to association studies comprising several ten SNPs, but can also be employed to analyze whole-genome data.

**Availability:** Software is available on request from the authors.

**Contact:** [robin.nunkesser@uni-dortmund.de](mailto:robin.nunkesser@uni-dortmund.de)

## 1 Introduction

Variations in the human genome can alter the risk of developing a disease. The by far most common type of such genetic variations are single nucleotide polymorphisms (SNPs) which occur when at a single base pair position different base alternatives exist. Since a SNP is typically biallelic, it can take three forms: A SNP is of the homozygous reference (or the homozygous variant) genotype if both chromosomes show the base that more (or less) frequently

---

\*to whom correspondence should be addressed

occur in the population, and it is of the heterozygous genotype if one of the bases is the less, and the other is the more frequent alternative.

One of the major goals of association studies is to identify SNPs and SNP interactions that lead to a higher disease risk. Since individual SNPs typically only have a slight to moderate effect – in particular, when considering complex diseases such as sporadic breast cancer – the focus is on the detection of interactions (Garte, 2001; Culverhouse *et al.*, 2002). The search for such interacting SNPs is additionally impeded by the facts that the interactions are usually of a high order, and that they are explanatory for relatively small subgroups of the patients (Pharoah *et al.*, 2004).

Various methods have been suggested for and applied to genotype data to identify SNP interactions. These procedures reach from exhaustive searches based on, e.g., multiple testing approaches (Marchini *et al.*, 2005; Boulesteix *et al.*, 2007; Goodman *et al.*, 2006; Ritchie *et al.*, 2001) to methods based on discrimination procedures (e.g., Lunetta *et al.*, 2004). For overviews on such approaches, see Heidema *et al.* (2006) and Hoh and Ott (2003).

One of the most promising methods is logic regression (Ruczinski *et al.*, 2003), an adaptive classification and regression procedure that tries to identify Boolean combinations of binary variables associated with the response (e.g., the case-control status). In several comparisons with other regression or discrimination approaches, logic regression has shown a good performance in its application to SNP data (Kooperberg *et al.*, 2001; Witte and Fijal, 2001; Ruczinski *et al.*, 2004; Schwender, 2007). Moreover, logic regression can be employed for detecting interactions and quantifying their importance (Kooperberg and Ruczinski, 2005; Schwender and Ickstadt, 2007).

For an application of logic regression to genotype data, each SNP needs to be coded by (at least) two dummy variables, as logic regression can only handle binary predictors, but SNPs can take three forms. Although this coding can be done in a biologically meaningful way (one dummy variable codes for a dominant effect, and the other for a recessive effect), it might be preferable to include each SNP as one variable in the analysis. Furthermore, the logic expressions generated by logic regression should be transformed into a disjunctive normal form (DNF) to identify the interactions, as the monomials included in the DNF can be interpreted as interactions.

Therefore, our procedure called GPAS (Genetic Programming for Association Studies) employs multi-valued logic, and attempts to detect DNFs associated with the response directly. To search for such DNFs, genetic programming (Koza, 1993) is used. Genetic programming naturally provides not a single best model, but a set of models (called individuals) that fit almost equally well, which is an advantage in the analysis of genotype data in which many competing models might exist.

In the following section, GPAS is introduced in detail. Afterwards, GPAS is applied to the genotype data from the GENICA study, a study dedicated to the identification of genetic and gene-environment interactions leading to a higher risk of developing sporadic breast cancer. In the analysis of this data set, GPAS is able to detect high-order SNP interactions associated with the case-control status. But GPAS is not restricted to association studies comprising several ten SNPs. It can also be used to analyze data from whole-genome studies. To exemplify this, GPAS is also applied to a subset of the HapMap data (The International HapMap Consortium, 2003).

## 2 Methods

We propose to use evolutionary algorithms - more precisely genetic programming (Koza, 1993) - for the analysis of genotype data.

In genetic programming, a set of *individuals* called *population* undergoes adaptations and afterwards a selection process based on *fitness* leading to a new *generation* of individuals. This procedure summarized in Algorithm 1 is iterated until a *termination criterion* is fulfilled.

---

**Algorithm 1 (Basic Genetic Programming Algorithm)**

1. Create an initial random population.
  2. Perform the following steps on the current generation:
    - (a) Select individuals in the population based on a selection scheme.
    - (b) Adapt the selected individuals.
    - (c) Evaluate the fitness value of the adapted individuals.
    - (d) Select individuals for the next generation according to a selection scheme.
  3. If the termination criterion is fulfilled, then output the final population. Otherwise, set the next generation as current and go to step 2.
- 

### 2.1 Genetic Programming for Association Studies

In the following, we customize the basic genetic programming algorithm presented in Algorithm 1 for our purpose, leading to our method GPAS.

#### Structure of the Individuals

In GPAS, multi-valued logic expressions in disjunctive normal form (DNF) are used as the structure for the individuals, where these logic expressions may exhibit any number of input states. In the application to SNP data, e.g., an input can take one of the following three states: 1 (coding for the homozygous reference), 2 (heterozygous), and 3 (homozygous variant).

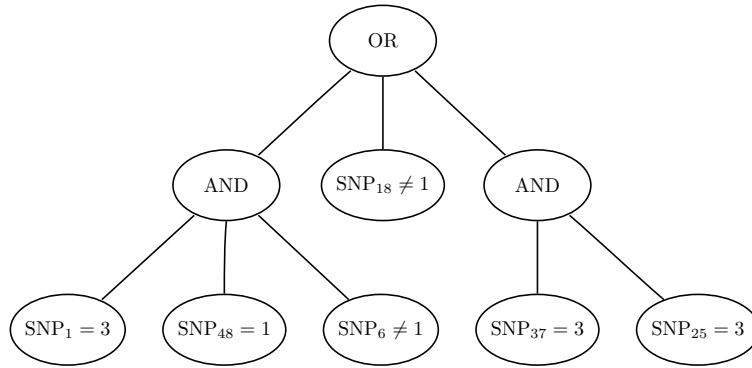
A logic expression in DNF is a disjunction of one or more monomials, where a monomial is a single literal or a conjunction of literals. Given, e.g., a set of variables  $X_1, \dots, X_m$ , each of which can take  $K$  values, the literals used in GPAS are

$$(X_i = k) \quad \text{and} \quad (X_i \neq k), \quad k = 1, \dots, K, \quad i = 1, \dots, m.$$

In Figure 1, an example of a generic tree representation of a logic expression  $L$  in DNF resulting from analyzing SNPs is shown, where

$$L = ((\text{SNP}_1 = 3) \wedge (\text{SNP}_{48} = 1) \wedge (\text{SNP}_6 \neq 1)) \\ \vee ((\text{SNP}_{18} \neq 1)) \vee ((\text{SNP}_{37} = 3) \wedge (\text{SNP}_{25} = 3)).$$

When used as a predictor in a case-control study, a patient would be classified as case if  $L$  is true, i.e. if all SNPs in at least one of the three monomials  $((\text{SNP}_1 = 3) \wedge (\text{SNP}_{48} =$



**FIGURE 1.** A logic expression in disjunctive normal form visualized as a tree.

$1) \wedge (\text{SNP}_6 \neq 1)$ ,  $(\text{SNP}_{18} \neq 1)$ , and  $((\text{SNP}_{37} = 3) \wedge (\text{SNP}_{25} = 3))$  show the genotypes indicated by the corresponding literals. Otherwise, the patient would be classified as control.

To store a logic expression in memory we use *trees* (see, e.g., [Cormen et al., 2001](#)) that are built according to the depicted tree representation as data structure. Using trees allows some very flexible and inexpensive operations: All of the adaptations described in the following are possible in amortized constant time when the children of a node in the tree are stored in a dynamic array.

## Operations for Adapt Individuals

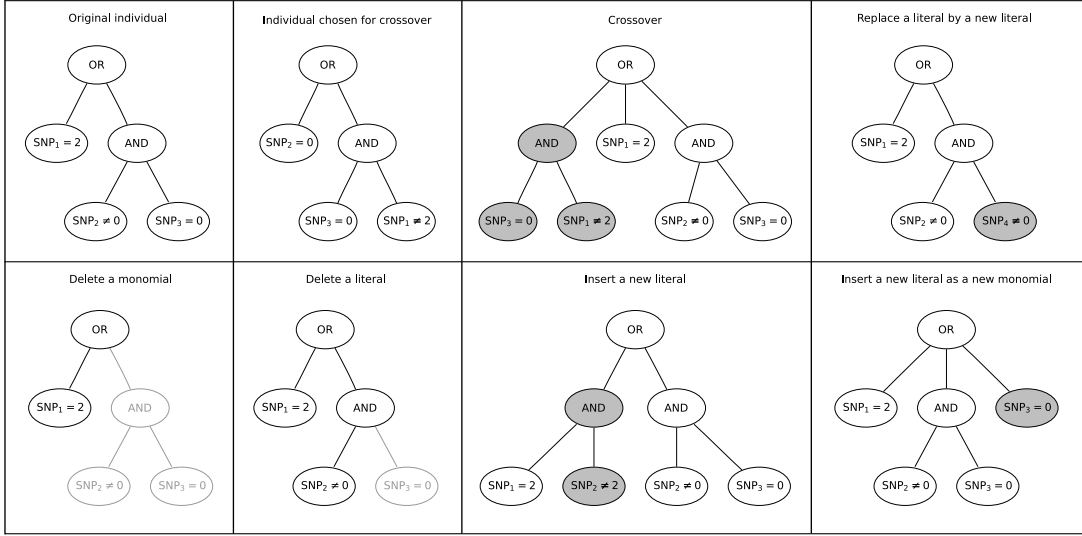
Initially, a population composed of two individuals, each consisting of one randomly selected literal, is created (corresponding to step 1 of Algorithm 1).

The set of candidate individuals for a new generation are constructed in steps 2a and 2b by selecting

- all individuals for *reproduction*, i.e. copying all individuals from the current generation,
- two individuals uniformly at random for *crossover*, i.e. combining one of the two individuals with one randomly chosen monomial from the other individual to create a new individual,
- five individuals uniformly at random for *mutation*, i.e. applying a random change to each of the individuals, where each of the possible mutations
  - inserting a new literal,
  - deleting a literal,
  - replacing a literal by a new literal,
  - inserting a new literal as a new monomial,
  - deleting a monomial.

is applied to exactly one of the five individuals.

In the latter adaptation, the literals or monomials that should be deleted are chosen uniformly at random, and the new literals are also selected at random and inserted into a randomly



**FIGURE 2.** Examples for the crossover and the different mutations used in GPAS.

chosen monomial or as a new monomial. An overview on the crossover and the mutations is given in Figure 2.

Note that the usage of crossover is discussed controversial (see e.g. [Banzhaf et al., 1998](#)). However, the crossover operation we propose does not disrupt the structure of the individuals and is therefore different from the criticized crossover operations. In the applications considered in this paper, it accelerates computation.

## Fitness and Selection

To determine which of the new and reproduced individuals are selected to be part of the next generation, we compute the fitness for each individual and select the best ones (corresponding to Step 2c and 2d of Algorithm 1, respectively).

We are interested in logic expressions that explain as many observations as possible, while being as short as possible. To achieve both goals equally well we conduct a multi-objective optimization by using multidimensional fitness values. The basic objectives of our optimization may be transferred to fitness values easily. Explaining as many observations as possible, for instance, translates to fitness values measuring the amount of data values fulfilled.

In the context of multi-objective optimization, an individual *dominates* another individual, if at least one component of its fitness value is superior, and none is inferior. An individual is *pareto optimal*, if it is not dominated by another individual.

Consequently, we seek to find pareto optimal individuals that offer a set of well fitting models.

For the new generation of individuals that is derived after the adaptations, we choose only individuals that are not dominated by other individuals. Thus, we conduct a *domination selection*. For our purposes, we use three objectives (see Sections 2.2 and 2.3), which leads to a bigger population and allows a more specialized search. For our two tasks – identification of interesting interactions and discrimination – we employ two slightly different fitness functions that are also described in Sections 2.2 and 2.3, respectively.

The major computational part of the fitness evaluation is to determine the number of cases and controls classified correctly by the logic expression.

For fast fitness computation, we additionally store a bitset in each node of the tree representing the logic expression. The bitset consists of as many bits as there are observations in the data set, and the  $i$ -th bit is true if the logic expression is true for the SNP forms of the  $i$ -th observation and false otherwise.

The bitsets of the literals are initially computed for all possible literals. If a monomial of the logical expression is changed during a mutation operation the bitset of the monomial is recomputed using the bitsets of its literals. The computation is sped up, since the bitsets of the other monomials remain unchanged and can be reused to compute the bitset of the whole logic expression. In addition, bitsets are compact and allow fast logic operations. For example, one logic operation of the bitset of the whole logic expression with the bitset describing the case-control status suffices to compute the number of cases and controls predicted correctly.

## Termination Criterion

We need termination criteria for the genetic programming process in order to derive a final population building the models (step 3 of Algorithm 1). Natural termination criteria used by GPAS are the excess of a certain number of generations or of a certain fitness value. Another possibility is to terminate the execution if the algorithm *stagnates*, i.e. no new individuals survived selection for a given number of generations.

## 2.2 Identification of Interactions

A major influence factor on the objective of an analysis is the choice of the fitness evaluation function. Interactions that explain subsets of the cases have to contradict with as few controls as possible. We, therefore, employ a fitness evaluation function that emphasizes this by including the number of correctly predicted controls in two of the objectives. The fitness of an individual is thus evaluated by the fitness function  $f_1$  that maps a logic expression to the following triple (corresponding to three objectives):

- (Maximize the) mean of the proportions of correctly classified cases and correctly classified controls.
- (Maximize the) number of controls the logic expression correctly predicts.
- (Minimize the) length of the logic expression, i.e. the number of literals of the logic expression.

A further modification of the general genetic algorithm is that we do not allow individuals to become too big. In the search for high-order interactions, we, furthermore, prohibit the algorithm from constructing polynomials, i.e. individuals, with more than two monomials.

To aid the detection of high-order interactions, we additionally devise a visualization of the resulting models. The interactions in the model are displayed in a tree showing many different interactions at a glance. To obtain this visualization (for an example of a resulting tree, see Figure 3), we proceed as follows:

1. Obtain the set  $\mathcal{M}$  of all monomials occurring in the resulting models.

2. Search for the most common literal  $\ell$  in  $\mathcal{M}$ , and determine the set  $\mathcal{M}_\ell$  of monomials containing  $\ell$ .
3. Exclude  $\ell$  from all monomials in  $\mathcal{M}_\ell$  to construct  $\mathcal{M}_{-\ell}$ .
4. Repeat steps 2-3 with  $\mathcal{M} := \mathcal{M}_{-\ell}$  and  $\mathcal{M} := \mathcal{M} \setminus \mathcal{M}_\ell$  until  $\mathcal{M} = \emptyset$ .

We additionally store information on how often the resulting interactions and partial interactions occur, and on how many observations they explain.

## 2.3 Discrimination

For discrimination, the first objective of  $f_1$  is replaced by

- (Maximize the) number of cases the logic expression predicts correctly.

leading to the fitness function  $f_2$ . Thus, predicting cases is treated in the same way as predicting controls. Additionally, we restrict the size of the individuals, but not the number of monomials comprised in an individual.

For class prediction of new observations, either the single best individual is used, or an ensemble of models is considered either by averaging over a set consisting of the best individuals, or by applying bagging (Breiman, 1996) to GPAS.

## 2.4 GPAS

To summarize, we propose the following specialized genetic programming algorithm called GPAS for the analysis of genotype data.

---

### Algorithm 2 (GPAS)

1. Create an initial random population composed of two individuals each of which consists of one randomly selected literal.
2. Perform the following steps on the current generation:
  - (a) Select all individuals in the population for reproduction, and draw seven of the individuals uniformly at random.
  - (b) Conduct each of the following adaptations to one (mutations) or two (crossover) of the seven randomly selected individuals.
    - Perform a crossover.
    - Insert a new literal.
    - Delete a literal.
    - Replace a literal by a new literal.
    - Insert a new literal as a new monomial.
    - Delete a monomial.
  - (c) Evaluate the fitness value of the adapted and reproduced individuals with fitness function  $f_1$  or  $f_2$ .



- (d) Select all adapted and reproduced individuals that are not dominated for the next generation.
  3. If the termination criterion is fulfilled, then output the final population. Otherwise, set the next generation as current and go to step 2.
- 

## 3 Data Sets

### 3.1 GENICA

The GENICA study is an age-matched and population-based case-control study carried out by the Interdisciplinary Study Group on **Gene ENvironment Interaction and Breast CAncer** in Germany (<http://www.genica.de>), a joint initiative of researchers dedicated to the identification of genetic and environmental factors associated with sporadic breast cancer. Cases and controls have been recruited in the greater Bonn, Germany, region. Apart from exogenous risk factors such as reproduction variables, hormone variables and life style factors, the genotypes of about 100 polymorphisms have been assessed from these women (for details on the GENICA study, see [Justenhoven \*et al.\*, 2004](#)).

In this paper, the focus is on a subset of the genotype data from the GENICA study. More precisely, data of 1,258 women (609 cases and 649 controls) and 63 SNPs are available for the analysis. Since a small number of observations show a large number of missing values, we remove all women with more than five missing values leading to a total of 1,191 observations (561 cases and 630 controls). The remaining missing values are replaced SNP-wise by random draws from the marginal distribution.

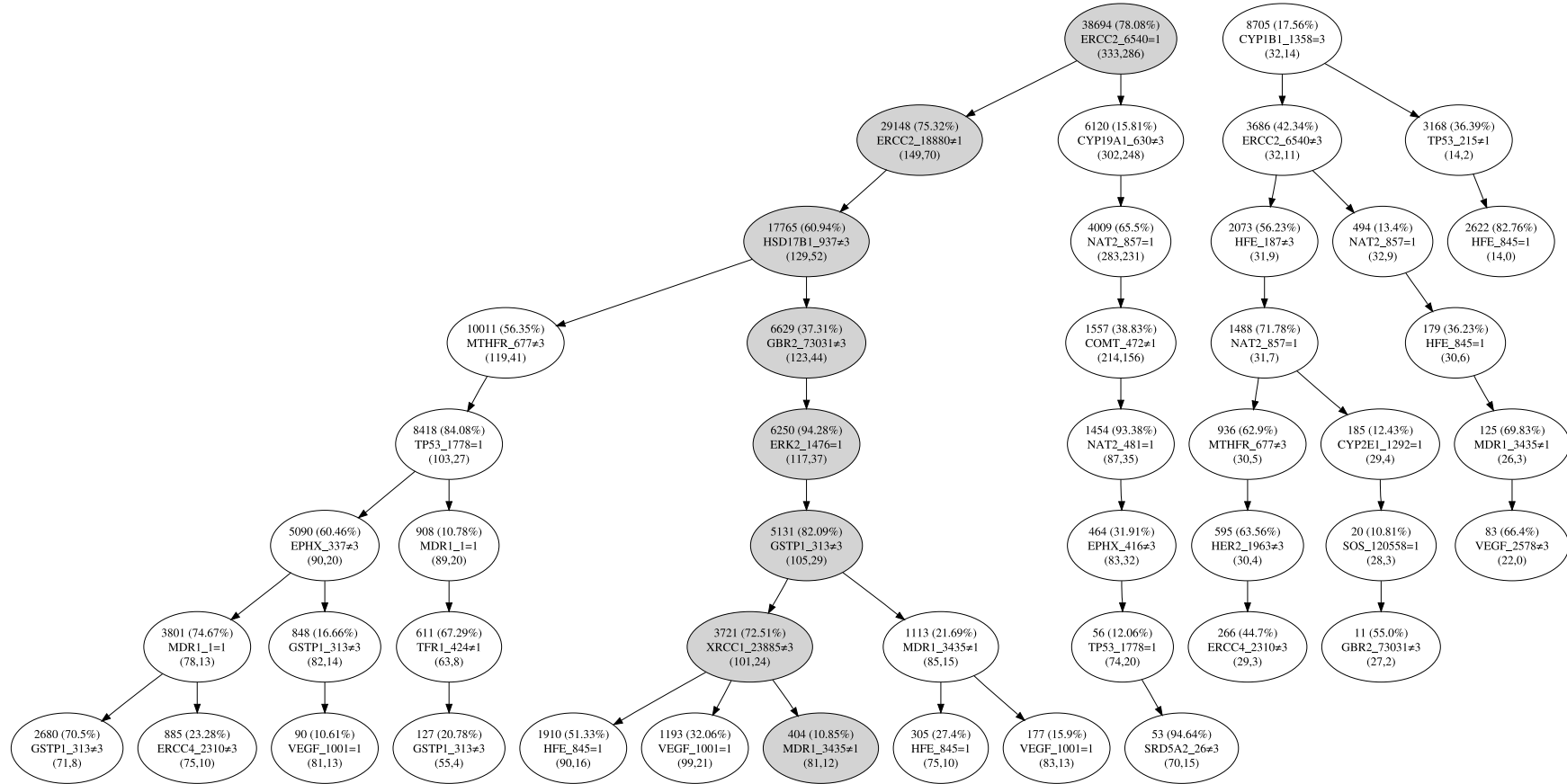
### 3.2 HapMap

The goals of the International HapMap Project ([The International HapMap Consortium, 2003](#); <http://www.hapmap.org>) are the development of a haplotype map of the human genome and the comparison of genetic variations of individuals from different populations. To achieve this goal, millions of SNPs have been genotyped for each of 270 people from four different populations.

In this paper, the SNP data of 45 unrelated Han Chinese from Beijing and 45 unrelated Japanese from Tokyo measured by employing the Affymetrix GeneChip Mapping 500K Array Set are considered.

This array set consists of two chips (the Nsp and the Sty array named after the restriction enzyme used on the respective chip) each enabling the genotyping of about 250,000 SNPs. Here, we focus on the BRLMM genotypes (Bayesian Robust Linear Model with Mahalanobis distance; [Affymetrix, 2006](#)) of the 262,264 SNPs from the Nsp array that can be downloaded from [http://www.affymetrix.com/support/technical/sample\\_data/500k\\_hapmap\\_genotype\\_data.affx](http://www.affymetrix.com/support/technical/sample_data/500k_hapmap_genotype_data.affx).

All SNPs showing one or more missing genotypes (54,400 SNPs), for which not all three genotypes are observed (75,481 SNPs), or that have a minor allele frequency less than or equal to 0.1 (10,609 SNPs) are excluded in this order from the analysis leading to a data set composed of the genotypes of 121,774 SNPs and 90 individuals.



**FIGURE 3.** Excerpt from the tree visualization of the models resulting from the application of GPAS to the GENICA data set. Each path from the root to an inner node or leaf represents an interaction occurring in the final population. The first line in each node consists of the number of monomials containing the corresponding interaction and the percentage of monomials consisting of the ancestral interaction that also contain the literal represented by the node, where this literal is displayed in the second line. The third line shows the number of cases and controls explained by the corresponding interaction.

## 4 Results

The following analyses are conducted on a Pentium 4 CPU with 2.56 GHz and 1024 MB of RAM.

### 4.1 Identification of Interesting SNP Interactions

In association studies concerned with sporadic breast cancer, it is assumed that not individual SNPs, but combinations of many SNPs have an high impact on the cancer risk, and that each of these interactions is a risk factor for a particular (relatively small) subgroup of patients (Pharoah *et al.*, 2004). In the analysis of the GENICA data set, we are thus interested in identifying high-order interactions explaining several ten cases, but only a few controls.

As mentioned in Section 2, we therefore constrain each individual in GPAS to consist of a maximum of two monomials. As  $\text{SNP}_i \neq 1$  codes for a dominant effect of  $\text{SNP}_i$ , and  $\text{SNP}_i = 3$  for a recessive effect, we restrict the set of literals used in GPAS to these two literals and their respective complements, i.e.  $\text{SNP}_i = 1$  and  $\text{SNP}_i \neq 3$ .

In this application of GPAS to the GENICA data set, we gather the individuals of 50 independent runs each of which stops after 500,000 generations, which takes about ten minutes. From the resulting 49,564 individuals, the tree visualization described in Section 2.2 is constructed. A pruned version of this tree is shown in Figure 3. For example, the eight literals marked by a gray background form an interaction that explains, i.e. a monomial that is true for, 81 cases and only 12 controls, and is contained in 404 of the individuals.

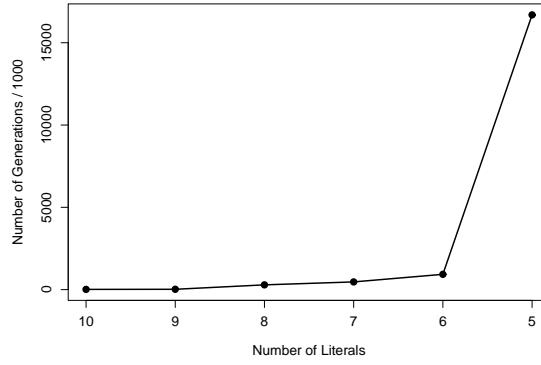
This figure also reveals that the interesting SNP interactions contain  $(\text{ERCC2\_6540} = 1) \wedge (\text{ERCC2\_18880} \neq 1)$ , i.e. an interaction of the two SNPs ERCC2\_6540 (refSNP ID: rs1799793) and ERCC2\_18880 (rs1052559) from the gene ERCC2 (Excision Repair Cross-Complementing group 2; formerly XPD), which itself explains 149 cases and 70 controls. This two-way interaction has already been found by Justenhoven *et al.* (2004) and by Schwender and Ickstadt (2007), but they were not able to identify interactions of higher orders with better odds ratios.

To examine if the exclusion of  $(\text{SNP}_i = 2)$  and  $(\text{SNP}_i \neq 2)$  has a large influence on the detection of interesting interactions, we also apply GPAS to the GENICA data set using the complete set of literals. In this analysis, some of the literals in the identified monomials are indeed of this type. However, these literals have mostly only a small effect, or they are equivalent to, e.g.,  $(\text{SNP}_i = 1)$ . For example, the interaction

$$\begin{aligned} & (\text{ERCC2\_6540} = 1) \wedge (\text{ERCC2\_18880} \neq 1) \\ & \wedge (\text{TFR1\_424} \neq 1) \wedge (\text{CYP1A1\_2452} = 1) \\ & \wedge (\text{MDR1\_1} \neq 2) \wedge (\text{TP53\_1778} \neq 2) \end{aligned}$$

detected in this application, which explains, i.e. is true for, 73 cases and 16 controls, contains two literals of the form  $(\text{SNP}_i = 2)$ . However,  $(\text{MDR1\_1} \neq 2)$  is actually  $(\text{MDR1\_1} = 1)$ , as none of the observations exhibit the homozygous variant genotype at this SNP, and replacing  $(\text{TP53\_1778} \neq 2)$  by  $(\text{TP53\_1778} = 1)$  would reduce the number of correctly predicted cases from 73 to 72, while the number of explained controls stays at 16.

To exemplify that GPAS is not restricted to data sets consisting of several ten to a few hundred SNPs, but can also be applied to data from whole genome studies, we apply GPAS to the subset of the HapMap data set described in Section 3.2. As it might be possible that



**FIGURE 4.** Number of generations (in thousands) in which individuals of certain lengths predicting all observations correctly are found in the application of GPAS to the HapMap data set.

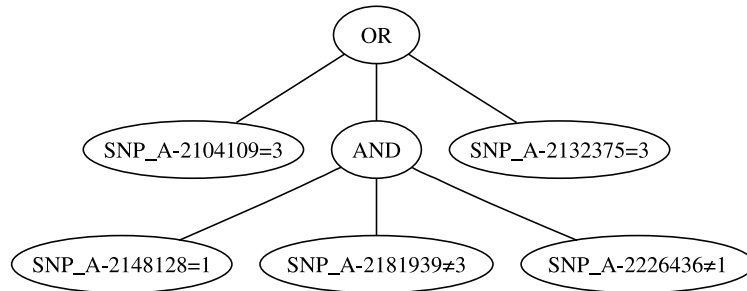
individual SNPs have a large influence in this example, we do not restrict the number of monomials in an individual. Furthermore, we only run GPAS once but without a termination criterion. All other settings remain unchanged compared to the analysis of the GENICA data set.

After running for nine minutes, GPAS detects an individual composed of ten literals in generation 13,683 that can be used to distinguish between the Japanese and the Han Chinese unambiguously: If at least one of the six monomials

$$\begin{aligned}
 &((\text{SNP\_A-1840639} = 1)), \\
 &((\text{SNP\_A-1862578} = 1)), \\
 &((\text{SNP\_A-1888933} = 3)), \\
 &((\text{SNP\_A-1983282} = 1) \wedge (\text{SNP\_A-2227333} = 3)), \\
 &((\text{SNP\_A-1849099} \neq 1) \wedge (\text{SNP\_A-2046537} \neq 1)), \\
 &((\text{SNP\_A-2030395} = 1) \wedge (\text{SNP\_A-1940113} \neq 1) \wedge (\text{SNP\_A-4200881} \neq 3))
 \end{aligned}$$

is true, then the person is from Japan (or more exactly, from Tokyo). Otherwise, it is a Han Chinese from Beijing.

This individual can still be optimized by reducing the number of SNPs (which is the third objective used in GPAS). Shortly after detecting this individual, GPAS finds individuals down



**FIGURE 5.** Individual composed of five SNPs that is identified by GPAS in the HapMap data set. It can be used to distinguish between Japanese and Han Chinese.

to length six (see Figure 4), and finally in generation 16,691,641 an individual composed of five literals/SNPs and displayed in Figure 5 is identified, where each of these individuals predict all observations correctly.

## 4.2 Discrimination

To examine how the misclassification rate depends on the number of variables in the model, GPAS is applied to the GENICA data set considering individuals composed of differing numbers of literals. For comparison, the GENICA data set is also analyzed using logic regression (Ruczinski *et al.*, 2003), where the number of variables allowed is constrained in the different applications. Since logic regression requires binary predictors, the  $i$ -th SNP,  $i = 1, \dots, m$ , is split into the two dummy variables

SNP <sub>$i1$</sub> : “SNP <sub>$i$</sub>  is not of the homozygous reference genotype.”

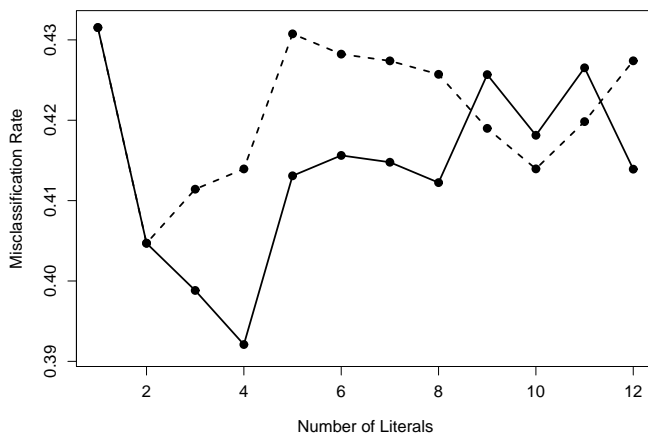
SNP <sub>$i2$</sub> : “SNP <sub>$i$</sub>  is of the homozygous variant genotype.”

where SNP <sub>$i1$</sub>  codes for a dominant effect of SNP <sub>$i$</sub> , and SNP <sub>$i2$</sub>  for a recessive effect. Note that SNP <sub>$i1$</sub> ,  $\overline{\text{SNP}}_{i1}$ , SNP <sub>$i2$</sub> , and  $\overline{\text{SNP}}_{i2}$  correspond to SNP <sub>$i$</sub>   $\neq$  1, SNP <sub>$i$</sub>  = 1, SNP <sub>$i$</sub>  = 3, and SNP <sub>$i$</sub>   $\neq$  3, respectively.

For each number of variables considered, we let GPAS run for 10,000 generations, which takes about one minute for each run.

In Figure 6, the resulting misclassification rates estimated by ten-fold cross-validation are displayed. This figure shows that the misclassification rates of both GPAS and logic regression are equal if the number of literals is less than 3. This is due to the fact that both use ((ERCC2\_6540 = 1)) or ((ERCC2\_6540 = 1)  $\wedge$  (ERCC2\_18880  $\neq$  1)), respectively, as classification rule in any of the respective iterations of the cross-validation. However, the misclassification rate of GPAS becomes smaller than the one of logic regression if the models are allowed to be composed of three to eight variables.

For a comparison of GPAS with further discrimination methods, CART (Breiman *et al.*, 1984), bagging (Breiman, 1996) and Random Forests (Breiman, 2001) are applied to the



**FIGURE 6.** Misclassification rates of GPAS (solid line) and logic regression (dashed line) in their applications to the GENICA data sets with restricted numbers of literals/variables in the individuals/models.

**TABLE 1.** Misclassification rates of the applications of several discrimination methods to both the GENICA and the HapMap data set.

	Logic			Random	
	GPAS	Regression	CART	Bagging	Forests
GENICA	0.392	0.405	0.429	0.457	0.450
HapMap	0.011	0.144	0.356	0.022	0.011

GENICA data set, where the parameters of the latter two procedures are optimized over several values. (In both bagging and Random Forests, different numbers of trees are considered. Additionally, different numbers of randomly chosen variables at each node are used in Random Forests.)

In Table 1, the misclassification rates of these applications are summarized. This table reveals that GPAS leads to less misclassifications than the other discrimination procedures.

For the application of these discrimination methods to the HapMap data set, the number of variables has to be reduced to a size that these approaches can handle. We therefore use the Significance Analysis of Microarrays (SAM; Tusher *et al.*, 2001) adapted for categorical data (Schwender, 2005) to reduce the number of SNPs from 121,774 to 157, where this subset of SNPs exhibits an estimated FDR (False Discovery Rate) of 0.069.

All discrimination methods are then applied to this subset of SNPs, and the misclassification is estimated by nine-fold cross-validation, where each of the nine subsets is composed of five randomly chosen Han Chinese and five randomly chosen Japanese.

Since for each of the training sets several models might exist that predict all training observations correctly, we use the bagging version of GPAS to stabilize the discrimination.

We also stop after 10,000 generations, which takes about twelve minutes for one training (consisting of 100 runs due to the use of bagging).

As Table 1 shows, both GPAS and Random Forests only misclassify one observation, whereas the discrimination methods that use a single model as classification rule, i.e. CART and logic regression, show a comparatively high misclassification rate.

## 5 Discussion

A major goal of association studies is the identification of SNPs and more importantly SNP interactions that lead to a higher risk of developing a disease. When considering complex diseases such as sporadic breast cancer, such interactions are typically of a high order and only explain relatively small subsets of the patients. Thus, approaches are needed that are able to detect these risk factors.

In this paper, we have presented a procedure based on genetic programming that can cope with this task. Genetic programming has the advantage that it is a general purpose method that can handle changing demands flexibly such as different fitness functions or size-constraints. In addition, the maintenance of candidate solutions is expedient for the multi-objective problems we tackle.

In the analysis of the GENICA data set, the presented method called GPAS identifies high-order interactions that explain sets of about 100 observations from which only a few are

controls. As the application to the 121,774 SNPs from the HapMap data set shows, GPAS can also be used to analyze whole-genome data. Moreover, GPAS is not restricted to feature selection, but can also be employed for classification, where it outperforms other tree-based discrimination methods in the applications to both the GENICA and the HapMap data set.

Although GPAS has been developed in the context of SNP data, it can also be applied to other types of categorical data, where the numbers of levels the variables can take might differ between variables.

Furthermore, the design of GPAS is flexible: By default, the set of literals is composed of all possible values for any of the variables and their corresponding complements. It is, however, possible to constrain this set of literals. For ordinal data,  $>$  and  $<$  can be used as operators additionally to or instead of  $=$  and  $\neq$ . Another possibility is to exclude any of the moves. For example, removing crossover from the move set might not worsen the results, but is likely to increase the computation time, as more generations have to be considered before the best solution is found.

Currently, the inputs, but not the output of GPAS can be multi-valued, as we are mainly interested in case-control studies. However, an extension of the two-class to the multi-class case is planned.

Another idea is to formulate – similar to logic regression – GPAS in a regression framework such that continuous responses, that are, e.g., of interest in QTL (Quantitative Trait Loci) analyses, can also be considered.

## Acknowledgement

Financial support of the Deutsche Forschungsgemeinschaft (SFB 475, “Reduction of Complexity in Multivariate Data Structures”) is gratefully acknowledged. The authors would also like to thank Roland Friend and all partners within the GENICA research network for their cooperation, and in particular Hermann M. Bolt, Hiltrud Brauch, Ute Hamann, Jan Hengstler, Christina Justenhoven, Sylvia Rabstein and Anne Spickenheuer for helpful discussions, and Melanie Schmidt for her help in implementing GPAS.

## References

- Affymetrix (2006). BRLMM: An improved genotype calling method for the GeneChip Human Mapping 500k array set. Technical report, Affymetrix, Santa Clara, CA.
- Banzhaf, W., Francone, F. D., Keller, R. E., and Nordin, P. (1998). *Genetic programming: an introduction: on the automatic evolution of computer programs and its applications*. Morgan Kaufmann Publishers Inc., San Francisco, CA, USA.
- Boulesteix, A. L., Strobl, C., Weidinger, S., Wichmann, H. E., and Wagenpfeil, S. (2007). Multiple testing for SNP-SNP interactions: A flexible asymptotic framework. Technical report, Sylvia Lawry Centre, Munich, Germany.
- Breiman, L. (1996). Bagging predictors. *Mach. Learn.*, **26**, 123–140.
- Breiman, L. (2001). Random Forests. *Mach. Learn.*, **45**, 5–32.

- Breiman, L., Friedman, J. H., Olshen, R. A., and Stone, C. J. (1984). *Classification and regression trees*. Wadsworth, Belmont, CA.
- Cormen, T. H., Leiserson, C. E., Rivest, R. L., and Stein, C. (2001). *Introduction to algorithms*. The MIT Press, Cambridge, Mass, second edition.
- Culverhouse, R., Suarez, B. K., Lin, J., and Reich, T. (2002). A perspective on epistasis: Limits of models displaying no main effect. *Am. J. Hum. Genet.*, **70**, 461–471.
- Garte, S. (2001). Metabolic susceptibility genes as cancer risk factors: Time for a reassessment? *Cancer Epidemiol. Biomarkers Prev.*, **10**, 1233–1237.
- Goodman, J. E., Mechanic, L. E., Luke, B. T., Ambbs, S., Chanock, S., and Harris, C. C. (2006). Exploring SNP-SNP interactions and colon cancer risk using polymorphism interaction analysis. *Int. J. Cancer*, **118**, 1790–1797.
- Heidema, G. A., Boer, J. M. A., Nagelkerke, N., Mariman, E. C. M., van de A, D. L., and Feskens, E. J. M. (2006). The challenge for genetic epidemiologists: How to analyze large numbers of SNPs in relation to complex diseases. *BioMed Genet.*, **7**(23).
- Hoh, J. and Ott, J. (2003). Mathematical multi-locus approaches to localizing complex human trait genes. *Nat. Rev. Genet.*, **4**, 701–709.
- Justenhoven, C., Hamann, U., Pesch, B., Harth, V., Rabstein, S., Baisch, C., Vollmert, C., Illig, T., Ko, Y., Brüning, T., and Brauch, H. (2004). ERCC2 genotypes and a corresponding haplotype are linked with breast cancer risk in a German population. *Cancer Epidemiology, Biomarkers and Prevention*, **13**, 2059–2064.
- Kooperberg, C. and Ruczinski, I. (2005). Identifying interacting SNPs using Monte Carlo logic regression. *Genet. Epidemiol.*, **28**, 157–170.
- Kooperberg, C., Ruczinski, I., LeBlanc, M., and Hsu, L. (2001). Sequence analysis using logic regression. *Genet. Epidemiol.*, **21**, 626–631.
- Koza, J. R. (1993). *Genetic programming – On the programming of computers by means of natural selection*. The MIT Press, Cambridge, Mass.
- Lunetta, K. L., Hayward, L. B., Segal, J., and van Eerdewegh, P. (2004). Screening large-scale association study data: exploiting interactions using random forests. *BMC Genet*, **10**(32).
- Marchini, J., Donnelly, P., and Cardon, R. C. (2005). Genome-wide strategies for detecting multiple loci that influence complex diseases. *Nat. Genet.*, **37**, 413–416.
- Pharoah, P. D., Dunning, A. M., Ponder, B. A., and Easton, D. F. (2004). Association studies for finding cancer-susceptibility genetic variants. *Nat. Rev. Cancer*, **4**, 850–860.
- Ritchie, M. D., Hahn, L. W., Roodi, N., Bailey, L. R., Dupont, W. D., Parl, F. F., and Moore, J. H. (2001). Multifactor-dimensionality reduction reveals high-order interactions among estrogen-metabolism genes in sporadic breast cancer. *Am. J. Hum. Genet.*, **69**, 138–147.
- Ruczinski, I., Kooperberg, C., and LeBlanc, M. (2003). Logic regression. *J. Comput. Graph. Stat.*, **12**, 475–511.



- Ruczinski, I., Kooperberg, C., and LeBlanc, M. (2004). Exploring interactions in high-dimensional genomic data: An overview of logic regression, with applications. *J. Mult. Anal.*, **90**, 178–195.
- Schwender, H. (2005). Modifying microarray analysis methods for categorical data – SAM and PAM for SNPs. In C. Weihs and W. Gaul, editors, *Classification – The Ubiquitous Challenge*, pages 370–377. Springer, Heidelberg.
- Schwender, H. (2007). *Statistical analysis of genotype and gene expression data*. Ph.D. thesis, Department of Statistics, University of Dortmund, Germany.
- Schwender, H. and Ickstadt, K. (2007). Identification of SNP interactions using logic regression. *Biostat.* to appear.
- The International HapMap Consortium (2003). The International HapMap Project. *Nature*, **426**, 789–796.
- Tusher, V., Tibshirani, R., and Chu, G. (2001). Significance analysis of microarrays applied to the ionizing radiation response. *Proc. Natl. Acad. Sci. USA*, **98**, 5116–5124.
- Witte, J. S. and Fijal, B. A. (2001). Introduction: Analysis of sequence data and population structure. *Genet. Epidemiol.*, **21**, 600–601.